

Enantioselective ring expansion of prolinols and ring-closing metathesis: formal synthesis of (–)-swainsonine

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Dedicated to Pr. Tietze on the occasion of his 65th birthday

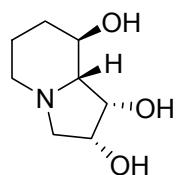
Abstract

An efficient enantioselective formal synthesis of (–)-swainsonine has been achieved in 14 steps with 14% global yield using an enantioselective ring enlargement of a substituted prolinol and a ring-closing metathesis as the key steps.

Keywords: Asymmetric synthesis, ring expansion, ring-closing metathesis, (–)-swainsonine

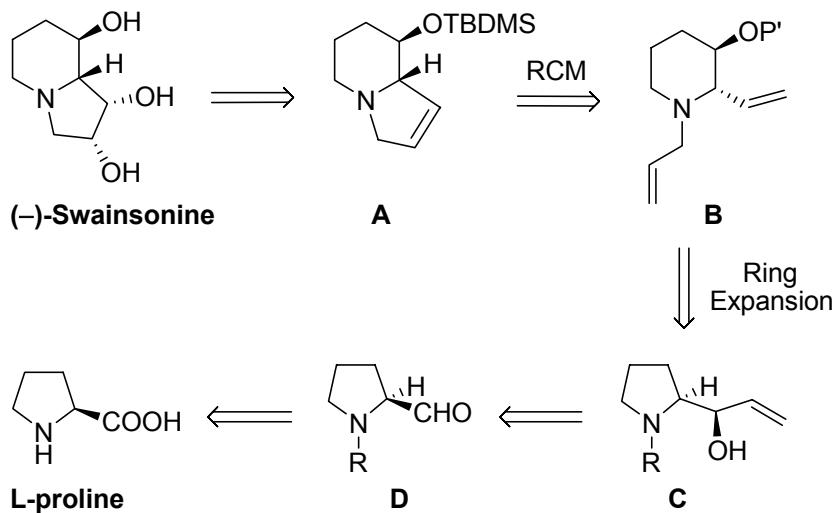
Introduction

The (1*S*,2*R*,8*R*,8*aR*)-1,2,8-trihydroxyindolizidine, (–)-swainsonine (Figure 1) belongs to the indolizidine class of alkaloid natural products. This compound has attracted the attention of synthetic chemists due to its interesting structure and potent biological properties. (–)-Swainsonine has been first isolated from the fungus *Rhizoctonia leguminicola* in 1973,¹ and has been later isolated from other plants² and fungus.³ (–)-Swainsonine has been the subject of many biological investigations and was found to be an effective inhibitor of both lysosomal α-mannosidase⁴ and mannosidase II.⁵ It has also demonstrated anticancer,⁶ anti-tumor proliferative⁷ and immunoregulating activity.⁸ Since the first total syntheses in 1984,^{9–12} there have been over 35 syntheses.^{13,14} In most of them, the starting material is a carbohydrate, which allows the introduction of the asymmetry and the control of the four asymmetric centers. By using D-erythrose, (–)-swainsonine could be synthesized in 8 steps, which is, to the best of our knowledge, the shortest synthesis up to now.¹⁵ A multigram scale synthesis in 10 steps from inexpensive D-ribose was also described recently.¹⁶

**Figure 1**

Except in the synthesis of swainsonine analogues, α -amino acids or derivatives have not been used to synthesize (-)-swainsonine. Here, we would like to report a formal synthesis of (-)-swainsonine starting from L-proline. The two key steps are an enantioselective ring enlargement of a substituted prolinol into a 3-hydroxypiperidine¹⁷ and a ring-closing metathesis¹⁸.

The synthesis of (-)-swainsonine was envisaged from the unsaturated bicyclic amino compound **A**, which would be synthesized by ring-closing metathesis applied to the amino diene **B**. This latter compound would be obtained by applying an enantioselective ring expansion to prolinol **C**, which will be synthesized by a diastereoselective addition of an organometallic species on prolinol **D**. This latter compound would be obtained from L-proline (Scheme 1).

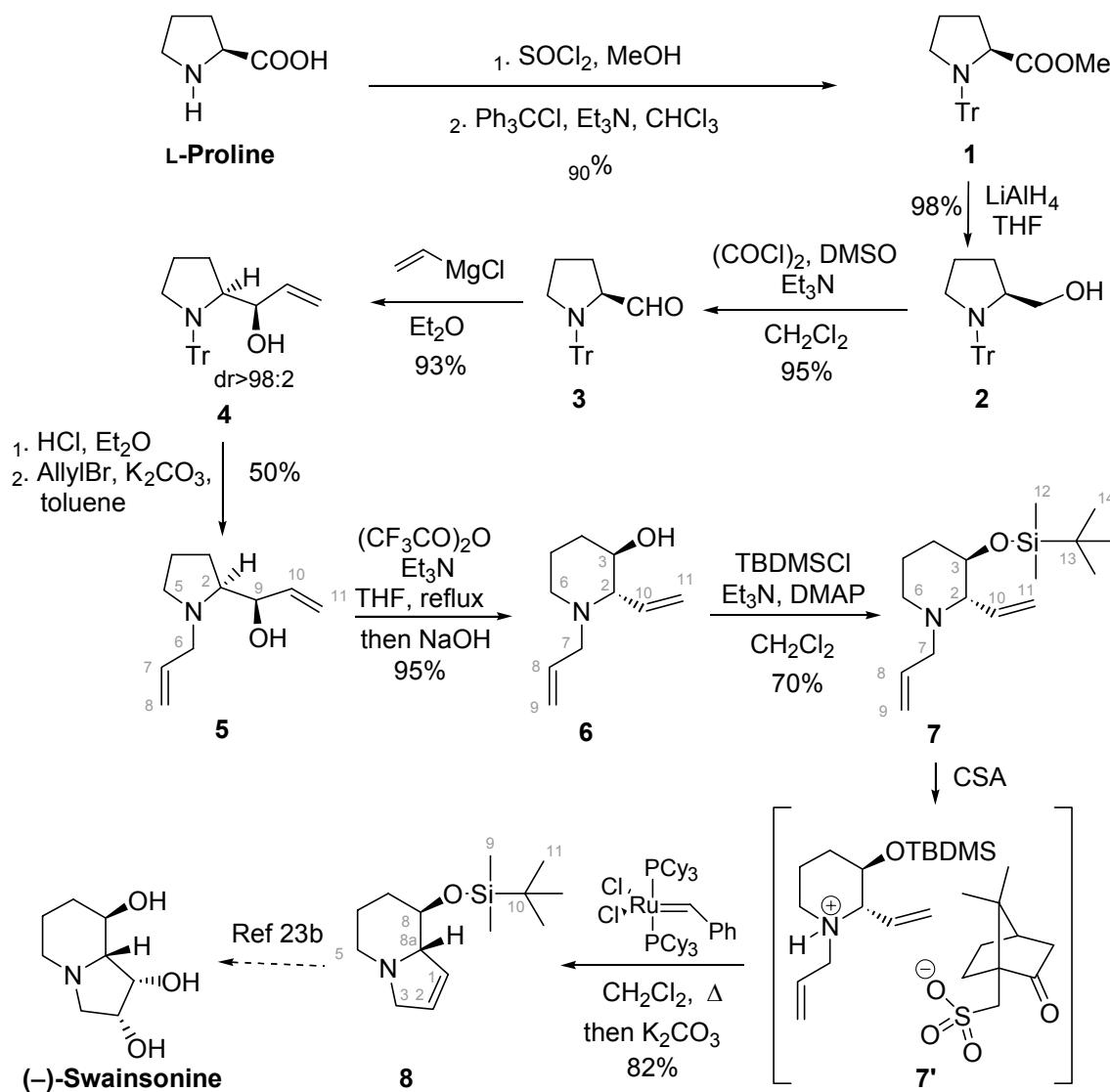
**Scheme 1**

Results and Discussion

The synthesis of (-)-swainsonine started from L-proline, which was transformed in five steps into prolinol **4**.¹⁹ After esterification (SOCl_2 , MeOH) and *N*-alkylation by using trityl chloride, amino ester **1** was isolated in 90% yield. This latter compound was converted into prolinol **2** in 98% yield by reduction using LiAlH_4 in THF. After oxidation under Swern conditions (oxalyl

chloride, DMSO, Et₃N, CH₂Cl₂), aldehyde **3** was isolated in 95% yield and treated with vinylmagnesium chloride in ether to give the allylic alcohol **4** as a single isomer (93% yield). We have to point out that the presence of the *N*-trityl group is of importance, as aldehydes of type **D** (R= alkyl, benzyl, Boc, Cbz, CO₂Me) are not configurationally stable in the presence of Grignard reagent, and the addition of organometallic onto these compounds usually led to a low diastereoselectivity.²⁰ On the contrary, *N*-tritylprolinal **3** is stable and addition of various nucleophiles is achieved with high level of diastereoselectivity.¹⁹ In order to build up the 5-membered ring of (–)-swainsonine by using a ring-closing metathesis, the *N*-trityl group in prolinol **4** was removed (HCl, Et₂O) and replaced by a *N*-allyl group (Allyl-Br, K₂CO₃, toluene) to produce the substituted prolinol **5** (50% yield), which is the precursor of the 3-hydroxypiperidine **6**. In agreement with previous results on the ring expansion of substituted prolinols,²¹ the enantioselective ring enlargement of prolinol **5** to piperidine **6** was performed by using trifluoroacetic anhydride (TFAA), Et₃N in THF and then NaOH. Under these conditions, 3-hydroxypiperidine **6** was isolated in 95% yield with a diastereomeric excess superior to 95%. After protection of the hydroxyl group in compound **6** (TBDMSCl, Et₃N, DMAP, CH₂Cl₂), piperidine **7** was isolated in 70% yield. As ruthenium catalysts are deactivated by amino groups, piperidine **7** was transformed to the ammonium salt **7'** by treatment with camphorsulfonic acid (CSA) and then treated with commercially available Grubbs catalyst first generation²² (5 mol. %) in refluxing CH₂Cl₂ and, after 6h, the reaction mixture was treated with K₂CO₃ to afford unsaturated bicyclic compound **8** in 82% yield (Scheme 2). The spectroscopic data of this latter compound are in agreement with those previously reported in the literature.²³ This approach constitutes a formal synthesis of (–)-swainsonine in 14 steps as the transformation of compound **8** into (–)-swainsonine was described in 4 steps in the literature.^{23b}

In conclusion, we have developed a short synthesis of (–)-swainsonine in 14% overall yield from L-proline by using a highly diastereoselective addition of vinyl Grignard reagent on *N*-tritylprolinal, an enantioselective ring expansion of a substituted prolinol into a 3-hydroxypiperidine to build up the 6-membered ring and a final ring-closing metathesis to construct the 5-membered ring. Further applications of this approach to swainsonine analogues and biological testing are ongoing.

**Scheme 2****Experimental Section**

General Procedures. Commercially available reagents and solvents were used as received. Anhydrous solvents were distilled. Tetrahydrofuran and diethyl ether were purified by distillation from sodium and benzophenone, methylene chloride was dried by distillation from CaH_2 . TLC was performed on Merck 60F₂₅₄ silica gel plates and visualized either with a UV lamp (254 nm), or by using a solution of $\text{KMnO}_4/\text{K}_2\text{CO}_3/\text{NaOH}$ in water followed by heating. Flash chromatography was performed with Merck Geduran Si60 silica gel (40–63 μm). Infrared (IR) spectra were recorded on a Bruker TENSOR™ 27 (IRFT); wavenumbers are indicated in cm^{-1} . $^1\text{H-NMR}$ spectra were recorded on a Bruker AVANCE 400 at 400 MHz and data are

reported as follows: chemical shift in ppm from tetramethylsilane as an internal standard, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), integration, assignment. ^{13}C -NMR spectra were recorded on a Bruker AVANCE 400 at 100 MHz and data are reported as follows: chemical shift in ppm from tetramethylsilane with the solvent as an internal indicator (CDCl_3 δ 77.0 ppm), multiplicity with respect to proton (deduced from DEPT experiments, s = quaternary C, d = CH, t = CH_2 , q = CH_3), assignment. Mass spectra with electronic impact (MS-EI) were recorded from a Hewlett-Packard tandem 5890A GC (12 m capillary column) – 5971 MS (70 eV). Mass spectra with chemical ionization (MS-CI) and high resolution mass spectra (HRMS) were performed by the Centre de Spectrochimie Organique de l'Ecole Normale Supérieure Ulm (Paris). Optical rotations were measured on a Perkin-Elmer 343 polarimeter in a 10 cm cell.

(R)-1-[(S)-1-Allyl-pyrrolidin-2-yl]-prop-2-en-1-ol (5). To a stirred solution of prolinol **4**¹⁹ (5.33 g, 14.4 mmol, 1.0 equiv) in Et_2O (40 mL) at 0 °C, was added HCl (5M aqueous solution, 29 mL) dropwise. The cooling bath was removed and after 24 h of vigorous stirring at RT, the two phases were separated and the aqueous phase was washed three times by Et_2O (3×30 mL). Toluene (150 mL) was added to the aqueous phase, the mixture was cooled down to 0 °C, and K_2CO_3 (26 g), then tetrabutylammonium bromide (0.93 g, 2.9 mmol, 0.2 equiv) and allyl bromide (3.75 mL, 43.3 mmol, 3.0 equiv) were added to the reaction mixture. After 24 h of vigorous stirring at RT, the two phases were separated. The aqueous phase was extracted twice with ethyl acetate (2×50 mL). The organic phases were dried over Na_2SO_4 and the solvents were evaporated to dryness *in vacuo*. The crude product was purified by flash chromatography on silica gel (cyclohexane/ Et_2O : 50/50). Compound **5** was obtained as a yellow oil (1.2 g, 7.2 mmol, 50% yield). ^1H -NMR (CDCl_3 , 400 MHz): δ 1.58–1.72 (3H, H_3 and H_4), 1.82 (m, 1H, H_3'), 2.30 (m, 1H, H_5), 2.53 (m, 1H, H_2), 2.92 (dd, J =13.7, 7.5 Hz, 1H, H_6), 3.15 (m, 1H, H_5'), 3.36 (bs, 1H, OH), 3.48 (dd, J =13.7, 5.2 Hz, 1H, H_6'), 4.24 (m, 1H, H_9), 5.08–5.24 (m, 3H, H_8 and H_{11}), 5.33 (ddd, J =17.1, 1.8, 1.8 Hz, 1H, H_{11}'), 5.77 (ddd, J =16.9, 10.7, 5.4 Hz, 1H, H_{10}), 5.89 (dddd, J =17.2, 10.0, 7.5, 5.5 Hz, 1H, H_7). ^{13}C -NMR (CDCl_3 , 100 MHz): δ 23.4 (t, C_4), 24.0 (t, C_3), 54.3 (t, C_5), 56.5 (t, C_6), 67.2 (d, C_2), 69.8 (d, C_9), 115.4 (t, C_{11}), 116.9 (t, C_8), 135.8 (d, C_7), 137.7 (d, C_{10}). IR (neat): 3405, 3075, 2960, 2795, 1642, 1604, 1444, 1419, 1351, 1280, 1205, 1150, 992, 921 cm^{-1} . MS (CI, CH_4): m/z (relative intensity): 168 (MH^+ , 100), 128 (8), 150 (12), 126 (4), 110 (47), 98 (3). $[\alpha]_D^{20} = -54.3$ (c 1.0, CHCl_3). HRMS: Found: m/z 168.1390. Calcd for $\text{C}_{10}\text{H}_{18}\text{NO}$: (MH^+) 168.1388.

(2S,3R)-1-Allyl-2-vinylpiperidin-3-ol (6). Trifluoroacetic anhydride (1.5 mL, 10.6 mmol, 1.5 equiv.) was added dropwise to a solution of prolinol **5** (1.2 g, 7.1 mmol, 1.0 equiv) in THF (60 mL) at 0 °C. After 1 h, Et_3N (3.0 mL, 21.2 mmol, 3.0 equiv) was added dropwise, and the reaction mixture was stirred for 20 min at 0 °C and then heated to reflux for 15 h. After addition of NaOH (2.5 M aqueous solution, 15 mL), the mixture was stirred for 2 h at RT and then extracted with EtOAc (3×30 mL), dried over Na_2SO_4 , and evaporated to dryness *in vacuo*. The residue was purified by flash column chromatography on silica gel (cyclohexane/ EtOAc : 80/20) to give piperidine **6** (1.1 g, 6.7 mmol, 95% yield) as a yellow oil. ^1H -NMR (CDCl_3 , 400 MHz): δ

1.30 (m, 1H, H₄), 1.56 (m, 1H, H₅), 1.71 (m, 1H, H_{5'}), 1.98–2.04 (2H, H_{4'} and H₆), 2.23 (bs, 1H, OH), 2.50 (dd, *J*= 8.4, 8.4 Hz, 1H, H₂), 2.76–2.90 (2H, H_{6'} and H₇), 3.31–3.39 (2H, H₃ and H_{7'}), 5.10–5.19 (2H, H₉), 5.31–5.37 (2H, H₁₁), 5.70–5.90 (2H, H₈ and H₁₀). ¹³C-NMR (CDCl₃, 100 MHz): δ 22.7 (t, C₅), 30.8 (t, C₄), 50.9 (t, C₆), 58.3 (t, C₇), 69.9 (d, C₃), 73.1 (d, C₂), 117.8 (t, C₉), 120.3 (t, C₁₁), 134.8 (d, C₈), 137.9 (d, C₁₀). IR (neat): 3415, 3076, 2936, 2861, 2793, 1643, 1441, 1419, 1261, 1090, 994, 916, 889 cm⁻¹. MS (EI, 70eV) *m/z* (relative intensity): 167 (M⁺, 9), 140 (9), 126 (100), 122 (16), 110 (17), 108 (21), 96 (14), 94 (18), 82 (17), 71 (16), 68 (18), 56 (14). [α]_D²⁰ = + 50.2 (*c* 1.03, CHCl₃). HRMS: Found: *m/z* 168.1384. Calcd for C₁₀H₁₈NO: (MH)⁺ 168.1388.

(2*S*,3*R*)-1-Allyl-3-(*tert*-butyldimethylsilyloxy)-2-vinylpiperidine (7). To a solution of piperidine **6** (0.670 g, 4.0 mmol, 1.0 equiv) in CH₂Cl₂ (50 mL) at RT, DMAP (0.049 g, 0.4 mmol, 0.1 equiv), Et₃N (1.13 mL, 8.0 mmol, 2.0 equiv) and TBDMSCl (1.20 g, 8.0 mmol, 2.0 equiv) were successively added. After 18 h at RT, the reaction mixture was quenched with a saturated aqueous NaHCO₃ solution until pH ~ 10. The aqueous layer was extracted with CH₂Cl₂ (2×30 mL) and the combined organic phases were dried over Na₂SO₄ and filtered. The solvent was removed *in vacuo*. The crude oil was purified by flash column chromatography on silica gel (cyclohexane/EtOAc: 90/10) to give **7** (0.788 g, 2.8 mmol, 70% yield) as a colorless oil. ¹H-NMR (CDCl₃, 400 MHz): δ -0.02 (s, 3H, H₁₂), 0.00 (s, 3H, H₁₂), 0.83 (s, 9H, H₁₄), 1.28 (m, 1H, H₄), 1.52 (m, 1H, H₅), 1.65 (m, 1H, H_{5'}), 1.88–1.95 (2H, H_{4'} and H₆), 2.42 (dd, *J*= 8.6, 8.6 Hz, 1H, H₂), 2.74 (dd, *J*= 13.9, 8.0 Hz, 1H, H₇), 2.88 (m, 1H, H_{6'}), 3.35–3.49 (2H, H₃ and H_{7'}), 5.07–5.14 (2H, H₉), 5.19 (dd, *J*= 4.4, 1.9 Hz, 1H, H₁₁), 5.22 (m, 1H, H_{11'}), 5.58 (m, 1H, H₁₀), 5.82 (dddd, *J*= 17.0, 10.3, 8.0, 5.3 Hz, 1H, H₈). ¹³C-NMR (CDCl₃, 100 MHz): δ -4.3 (q, C₁₂), -4.2 (q, C₁₂), 18.1 (s, C₁₃), 23.4 (t, C₅), 25.9 (q, 3C₁₄), 34.0 (t, C₄), 51.6 (t, C₆), 58.2 (t, C₇), 71.8 (d, C₂), 73.7 (d, C₃), 117.6 (t, C₉), 119.4 (t, C₁₁), 135.1 (d, C₈), 138.9 (d, C₁₀). IR (neat): 2928, 2857, 2790, 1643, 1462, 1361, 1253, 1101, 997, 916, 834, 772 cm⁻¹. MS (EI, 70eV) *m/z* (relative intensity): 281 (M⁺, 4), 241 (20), 240 (100), 224 (15), 185 (9), 150 (10), 122 (10), 110 (56), 108 (23), 73 (28). [α]_D²⁰ = + 64.2 (*c* 0.67, CHCl₃). HRMS: Found: *m/z* 282.2257. Calcd for C₁₆H₃₂NOSi: (MH)⁺ 282.2253.

(8*R*,8a*S*)-8-(*tert*-Butyldimethylsilyloxy)-3,5,6,7,8,8a-hexahydroindolizine (8).^{23b} To a solution of piperidine **7** (79 mg, 0.28 mmol, 1.0 equiv) in CH₂Cl₂ (10 mL) at 0 °C, (+)-camphorsulfonic acid (72 mg, 0.31 mmol, 1.1 equiv) was added. After 10 min at 0 °C, the cooling bath was removed and Grubbs catalyst first generation was added by portions (3×8 mg, 0.035 mmol, 12.5 mol. %), each addition was followed by 2 h reflux. The reaction mixture was then treated with K₂CO₃ and filtered. The solvent was removed *in vacuo*. The crude oil was purified by flash column chromatography on silica gel (chloroform/MeOH: 90/10) to give **8** (58 mg, 0.23 mmol, 82% yield) as a yellow oil. ¹H-NMR (C₆D₆, 400 MHz): δ 0.05 (s, 3H, H₉), 0.07 (s, 3H, H₉), 0.99 (s, 9H, H₁₁), 1.27 (m, 1H), 1.41 (m, 1H), 1.59 (m, 1H), 1.89 (ddd, *J*= 12.2, 7.9, 4.0 Hz, 1H), 2.25 (ddd, *J*= 11.6, 11.6, 2.9 Hz, 1H, H₅), 2.72 (dd, *J*= 11.2, 4.8 Hz, 1H, H_{8a}), 3.04 (m, 1H, H_{5'}), 3.13 (dddd, *J*= 12.5, 6.5, 2.0, 2.0 Hz, 1H, H₃), 3.54 (dddd, *J*= 12.5, 3.9, 1.9, 1.9 Hz, 1H, H_{3'}), 3.60 (ddd, *J*= 10.2, 8.8, 4.4 Hz, 1H, H₈), 5.69 (ddd, *J*= 6.2, 4.0, 2.1 Hz, 1H, H₁),

6.18 (m, 1H, H₂). ¹³C-NMR (C₆D₆, 100 MHz): δ -4.5 (q, C₉), -4.1 (q, C₉), 18.3 (s, C₁₀), 25.2 (t, C₆), 26.1 (q, 3C₁₁), 35.1 (t, C₇), 49.1 (t, C₅), 58.3 (t, C₃), 72.5 (d, C₈), 74.8 (d, C_{8a}), 128.9 (d, C₁), 131.5 (d, C₂). IR (neat): 2931, 2886, 2857, 2778, 1467, 1368, 1253, 1148, 1095, 839, 775 cm⁻¹. MS (Cl, CH₄) m/z (relative intensity): 254 (MH⁺, 100), 144 (3), 120 (24), 111 (3). [α]_D²⁰ = -88.9 (c 0.70, C₆H₆).

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References

1. Guengerich, F. P.; DiMari, S. J.; Broquist, H. P. *J. Am. Chem. Soc.* **1973**, *95*, 2055.
2. (a) Colegate, S. M.; Dorling, P. R.; Huxtable, C. R. *Aust. J. Chem.* **1979**, *32*, 2257. (b) Ermayanti, T. M.; McComb, J. A.; O'Brien, P. A. *Phytochemistry* **1994**, *36*, 313.
3. Tamerler, C.; Kesharaz, T. *Biotechnol. Lett.* **1999**, *21*, 501.
4. Liao, Y. F.; Lal, A.; Moreman, K. W. *J. Biol. Chem.* **1996**, *271*, 28348.
5. (a) Elbein, A. D.; Solf, R.; Dorling, P. R.; Vosbeck, K. *Proc. Natl. Acad. Sci. U.S.A.* **1981**, *78*, 7393. (b) Kaushal, G. P.; Szumilo, T.; Pastuszak, I.; Elbein, A. D. *Biochemistry* **1990**, *29*, 2168. (c) Pastuszak, I.; Kaushal, G. P.; Wall, K. A.; Pan, Y. T.; Sturm, A.; Elbein, A. D. *Glycobiology* **1990**, *1*, 71.
6. (a) Goss, P. E.; Baker, M. A.; Carver, J. P.; Dennis, J. W. *Clin. Cancer Res.* **1995**, *1*, 935. (b) Goss, P. E.; Reid, C. L.; Bailey, D.; Dennis, J. W. *Clin. Cancer Res.* **1997**, *3*, 1077.
7. Dennis, J. W. *Cancer Res.* **1986**, *46*, 5131.
8. White, S. L.; Schweitzer, K.; Humphries, M. J.; Olden, K. *Biochem. Biophys. Res. Commun.* **1988**, *150*, 615.
9. Mezher, H. A.; Hough, L.; Richardson, A. C. *J. Chem. Soc., Chem. Commun.* **1984**, 447.
10. Fleet, G. W. J.; Gouh, M. J.; Smith, P. W. *Tetrahedron Lett.* **1984**, *25*, 1853.
11. Suami, T.; Tadano, K.; Iimura, Y. *Chem. Lett.* **1984**, 513.
12. Adams, C. E.; Walker, F. J.; Sharpless, K. B. *J. Org. Chem.* **1985**, *50*, 420.
13. For review see: (a) Nemr, A. E. *Tetrahedron* **2000**, *56*, 8579. (b) Pyne, S. G. *Current Organic Synthesis* **2005**, *2*, 39.
14. Guo, H.; O'Doherty, G. A. *Org. Lett.* **2006**, *8*, 1609.
15. Bennett, R. B.; Choi, J. R.; Montgomery, W. D.; Cha, J. K. *J. Am. Chem. Soc.* **1989**, *111*, 2580.
16. Pearson, W. H.; Ren, Y.; Powers, J. D. *Heterocycles* **2002**, *58*, 421.

17. For comprehensive reviews, see: (a) Cossy, J.; Gomez Pardo, D. *Chemtracts* **2002**, *15*, 579. (b) Cossy, J.; Gomez Pardo, D. *Targets in Heterocyclic Systems* **2002**, *6*, 1.
18. For reviews, see: (a) Fürstner, A. *Angew. Chem., Int. Ed. Engl.* **2000**, *39*, 3012. (b) Grubbs R. G.; Chang, S. *Tetrahedron* **1998**, *54*, 4413. (c) Armstrong, S. *J. Chem. Soc., Perkin Trans. I* **1998**, *371*. (d) Schuster, M. ; Blechert, S. *Angew. Chem., Int. Ed. Engl.* **1997**, *36*, 2036.
19. Bejjani, J.; Chemla, F.; Audouin, M. *J. Org. Chem.* **2003**, *68*, 9747.
20. (a) Andrès, J. M.; Pedrosa, R.; Pérez, A.; Pérez-Encabo, A. *Tetrahedron* **2001**, *57*, 8521. (b) Ma, D.; Pan, Q.; Han, F. *Tetrahedron Lett.* **2002**, *43*, 9401.
21. (a) Cossy, J.; Dumas, C.; Michel, P.; Gomez Pardo, D. *Tetrahedron Lett.* **1995**, *36*, 549. (b) Cossy, J.; Dumas, C.; Gomez Pardo, D. *Synlett* **1997**, *905*. (c) Cossy, J.; Dumas, C.; Gomez Pardo, D. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 1343. (d) Wilken, J.; Kossenjans, M.; Saak, W.; Haase, D.; Pohl, S.; Martens, J. *Liebigs Ann.* **1997**, *573*. (e) Langlois, N.; Calvez, O. *Synth. Commun.* **1998**, *28*, 4471. (f) Davis, P. W.; Osgood, S. A.; Hébert, N.; Sprankle, K. G.; Swayze, E. E. *Biotechnol. Bioeng.* **1999**, *61*, 143. (g) Cossy, J.; Dumas, C.; Gomez Pardo, D. *Eur. J. Org. Chem.* **1999**, *1693*. (h) Michel, P.; Rassat, A. *J. Org. Chem.* **2000**, *65*, 2572. (i) Cossy, J.; Mirguet, O.; Gomez Pardo, D. *Synlett* **2001**, *1575*. (j) Brandi, A.; Cicchi, S.; Paschetta, V.; Gomez Pardo, D.; Cossy, J. *Tetrahedron Lett.* **2002**, *43*, 9357. (k) Deyine, A.; Delcroix, J.-M.; Langlois, N. *Heterocycles* **2004**, *64*, 207. (l) Déchamps, I.; Gomez Pardo, D.; Karoyan, P.; Cossy, J. *Synlett* **2005**, *1170*. (m) Roudeau, R.; Gomez Pardo, D.; Cossy, J. *Tetrahedron* **2006**, *62*, 2388. (n) Mena, M.; Bonjoch, J.; Gomez Pardo, D.; Cossy, J. *J. Org. Chem.* **2006**, *71*, 5930.
22. Schwab, P.; France, M. B.; Ziller, J. W.; Grubbs, R. H. *Angew. Chem., Int. Ed. Engl.* **1995**, *34*, 2039.
23. (a) Mukai, C.; Sugimoto, Y.-I.; Miyazawa, K.; Yamaguchi, S.; Hanaoka, M.; *J. Org. Chem.* **1998**, *63*, 6281. (b) Buschmann, N.; Rückert, A.; Blechert, S. *J. Org. Chem.* **2002**, *67*, 4325.